McCampbell Analytical, Inc.
"When Quality Counts"
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UCMR 4 Sampling Instructions

UCMR 4 samples must not be composited (i.e., combined, mixed or blended); samples from each sampling point must be collected, preserved and processed separately.

PWSs that ship the samples for analysis should recognize that samples must be collected early enough in the day to allow adequate time to send these samples for overnight delivery to the laboratory. The PWS should avoid collecting samples on Thursday and should not collect samples on Friday, Saturday, or Sunday unless special arrangements have been made for the receipt of samples at the laboratory within 48-hours of collection.

Supplies should be assembled and taken to the sampling point at the same time. These will include:

- Cooler
- Sampling Containers
- Gloves
- Chain of Custody
- Labels
- Markers
- Goggles
- Bagged Ice
- Tape for securing the cooler

Optional:

- Paper Towels for drying the outside of the containers after sampling
- Plastic storage bags for ice or samples

General Sampling Procedure for All Methods:

Use the proper container to collect your sample and do not rinse out any preservatives present. Specific sample containers are required for all UCMR4 analyses to comply with each method. Samples will be rejected if collected in the incorrect container and will require re-sampling. If you have any questions, please contact us before sampling at 877-252-9262.

Wash your hands before and after sampling. Label sample bottles prior to sampling using a permanent marker. Label should include sample location, date and time collected.

Open the tap and allow the system to flush until the water temperature has stabilized (approximately 3 minutes). Decrease the flow to approximately ¼ inch across (width of a pencil). Collect samples from the flowing system.

Fill sample bottle at an angle pointing away from you. Fill to the shoulder, but do not flush out the preservation reagents. Cap the bottle and agitate by hand until preservative is dissolved.

Refrigerate samples or place samples on ice immediately after sampling prior to shipping to lab. Samples must be submitted to the lab within 48 hours of sampling. UCMR4 protocols have specific preservation and

temperature requirements therefore samples must arrive at the laboratory cold (<10°C) and in the correct containers or samples will be rejected.

Before submitting your samples to the lab, complete a chain of custody form for each point (EP).

EPA 200.8 Metals in Drinking Water

Bottles: 250 mL HDPE plastic (one bottle for each entry point)

Fill the bottle under a slow stream of water until the bottle is filled up to the neck.

EPA 525.3 Pesticides

Bottles: 1 L amber glass (three bottles for each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 530 Semi volatiles

Bottles: 1L amber glass (two bottles for each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 541 Alcohols

Bottles: 60 mL amber glass vials (two per each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 552.3: Haloacetic Acids

Bottles: 60 mL amber glass vials (two for each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 544: Microcystins & Nodularin

Bottles: 550 mL amber glass (three bottles for each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

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EPA 545: Cylindrospermopsin & Anatoxin-a

Bottles: 125 mL amber glass (one bottle for each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 546 (ELISA): Total Microcystins

Bottles: 125 mL amber glass (one bottle for each entry point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

Sample Transport and Storage

Samples should arrive at the laboratory packed on ice or with frozen gel packs in a sample bottle constructed of appropriate material as described in the analytical method. Samples that are shipped from the PWS should be sent by priority overnight delivery to the laboratory.

UCMR samples that arrive at the laboratory within 48 hours of collection must not exceed 10ºC.

Samples with water temperatures significantly above 10ºC at the time of collection (e.g., summer sampling events with elevated ambient water temperature), may be iced or refrigerated for a period of time prior to shipping to ensure the laboratory receipt temperature criteria are met.

Samples that arrive at the laboratory on the same day of sample collection (exclusively due to the close proximity of a PWS to the laboratory), may not yet have stabilized to 10ºC or less when they arrive at the lab. These samples are acceptable ONLY if packed on ice or with frozen gel packs immediately after sample collection and hence, delivered while samples are in the process of reaching an equilibrium temperature less than 10ºC. Samples must not be analyzed if they were not shipped properly.

For additional questions / clarification please contact:

Email: main@mccampbell.com
Phone: 877-252-9262
Sales – New client setup, Clarification of testing, pricing / TAT's
Login – Clarification for submitting samples